Peripheral Markers of Vascular Endothelial Dysfunction Show Independent but Additive Relationships with Brain-Based Biomarkers in Association with Functional Impairment in Alzheimer's Disease

⁶ Jonathan D. Drake^{a,b}, Alison B. Chambers^c, Brian R. Ott^{a,b}, Lori A. Daiello^{a,b} and for the

⁷ Alzheimer's Disease Neuroimaging Initiative¹

⁸ ^aAlzheimer's Disease and Memory Disorders Center, Rhode Island Hospital, Providence, RI, USA

^bDepartment of Neurology, Brown University Warren Alpert Medical School, Providence RI, USA

^cDepartment of Medicine, Brown University Warren Alpert Medical School, Providence RI, USA

Handling Associate Editor: Ruth Peters

11 Accepted 8 February 2021 Pre-press 10 March 2021

14 Abstract.

Background: Cerebrovascular dysfunction confers risk for functional decline in Alzheimer's disease (AD), yet the clinical interplay of these two pathogenic processes is not well understood.

- Objective: We utilized Alzheimer's Disease Neuroimaging Initiative (ADNI) data to examine associations between periph-
- erally derived soluble cell adhesion molecules (CAMs) and clinical diagnostic indicators of AD.
- 19 Methods: Using generalized linear regression models, we examined cross-sectional relationships of soluble plasma vascular
- cell adhesion molecule-1 (VCAM-1), intercellular adhesion molecule-1 (ICAM-1), and E-Selectin to baseline diagnosis and
- functional impairment (clinical dementia rating sum-of-boxes, CDR-SB) in the ADNI cohort (n=112 AD, n=396 mild
- cognitive impairment (MCI), n = 58 cognitively normal). We further analyzed associations of these biomarkers with brain-
- ²³ based AD biomarkers in a subset with available cerebrospinal fluid (CSF) data (*n* = 351). *p*-values derived from main effects

and interaction terms from the linear regressions were used to assess the relationship between independent and dependent

- variables for significance (significance level was set at 0.05 *a priori* for all analysis).
- **Results:** Higher mean VCAM-1 (p = 0.0026) and ICAM-1 (p = 0.0189) levels were found in AD versus MCI groups; however,
- not in MCI versus cognitively normal groups. Only VCAM-1 was linked with CDR-SB scores (p = 0.0157), and APOE $\varepsilon 4$
- ³⁰ genotype modified this effect. We observed independent, additive associations when VCAM-1 and CSF amyloid- β (A β ₄₂), total tau, phosphorylated tau (P-tau), or P-tau/A β ₄₂ (all < p = 0.01) were combined in a CDR-SB model; ICAM-1 showed a similar pattern, but to a lesser extent.

¹Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (http://adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can

ISSN 1387-2877 © 2021 – The authors. Published by IOS Press. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (CC BY-NC 4.0).

be found at: http://adni.loni.usc.edu/wp-content/uploads/how_ to_apply/ADNI_Acknowledgement_List.pdf

^{*}Correspondence to: Jonathan D. Drake, MD, Alzheimer's Disease and Memory Disorders Center, Rhode Island Hospital, 593 Eddy Street, Providence, RI 02903, USA. Tel.: +1 401 444 5745; Fax: +1 401 606 8555; E-mail: jdrake@lifespan.org.

33 Keywords: Alzheimer's disease, E-Selectin, intercellular adhesion molecule-1, vascular cell adhesion molecule-1

31 INTRODUCTION

Although amyloid deposition is considered to be 32 the central inciting event in the development of clin-33 ical Alzheimer's disease (AD) [1], age-associated 34 cerebral vascular disease and AD have long been rec-35 ognized as frequently co-morbid entities. Abundant 36 contemporary epidemiologic data point toward clin-37 ically relevant interrelationships between vascular 38 dysfunction and AD, as evidenced by a strong asso-39 ciations between mid-life vascular risk factors and 40 later-life clinical AD [2-9]. Mid-life vascular risk fac-41 tors have moreover been linked to later-life amyloid 42 burden [10, 11], including in those with high poly-43 genic risk for AD [10]. In support of this narrative, 44 declining dementia incidence in developed countries 45 is thought to be related to general improvements in 46 preventive disease management [12]. 47

Despite these findings and consistent neuropatho-48 logical observations of highly prevalent mixed 49 pathology in the elderly [13], the question of whether 50 concurrent cerebral vascular disease and AD patholo-51 gies represent biologically intertwined or parallel but 52 non-synergistic processes is difficult to fully eluci-53 date and has been a topic of frequent historical debate 54 [13-24]. Given the critically unmet need for effec-55 tive preventive and therapeutic strategies for AD and 56 the modifiable nature of vascular risk factors, disen-57 tangling various pathogenic processes that underlie 58 risk for age-related cognitive decline and identifica-59 tion of biomarkers that may be sensitive to modifiable 60 vascular-mediated risk is of high clinical import. 61

The majority of prior studies analyzing con-62 tributions of co-existing cerebrovascular and AD 63 pathology have found independent and/or additive 64 rather than interactive associations to risk for cog-65 nitive decline; however, most studies to-date have 66 largely focused on relationships of imaging-based 67 white matter disease burden to amyloid pathology 68 [23, 25, 26]. In contrast, results from more recent 69 studies using composite [27] or expanded [28, 29] 70 cerebrovascular biomarker panels or AD biomarkers 71 that are inclusive of amyloid, tau, and neurode-72 generative pathologies [26, 30] suggest that some 73

synergistic aspects of vascular-mediated associations to AD pathogenesis may have been previously underappreciated.

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

We sought to utilize peripherally derived molecular measures of vascular endothelial dysfunction as a means by which to assess for potentially more nuanced contributions of vascular risk to AD-associated pathologies and clinical AD. Cell adhesion molecules (CAMs) are expressed on the vascular endothelial surface and facilitate blood-borne leukocyte recruitment and trafficking across the vascular endothelium to sites of tissue damage via stereotyped stages: initial leukocyte tethering and rolling along the endothelial vessel wall (L-Selectin, P-Selectin, E-Selectin, and vascular cell adhesion molecule-1 (VCAM-1)), followed by firm adhesion (intercellular adhesion molecule-1 (ICAM-1), VCAM-1), and eventual chemotaxis and transendothelial cell migration (PECAM; Platelet And Endothelial Cell Adhesion Molecule) [31–36]. Membrane-bound CAMs are subsequently shed as soluble forms after a time delay, presumably as part of a feedback mechanism [37], and the soluble fraction can be conveniently measured in blood samples to detect processes of active inflammation involving the vascular endothelium.

Elevated circulating and/or cerebrospinal fluid (CSF) CAMs have been reported in relation to a variety of vascular risk factors and cardiovascular disease [38-44], cerebrovascular disease [45], neuroimmunological disorders such as multiple sclerosis [46, 47], and a broad spectrum of immune-mediated disorders including lupus erythematosus, rheumatoid arthritis, asthma, and cancer [39]. There have been a limited number of studies to-date assessing relationships of blood- or CSF-borne CAMs to cognitive aging and dementia in humans [32, 48, 57-62, 49-56]. However, samples sizes of bloodborne CAMs in most studies have been relatively small, and to our knowledge no studies thus far have tested for interactive associations between soluble plasma CAMs and CSF AD biomarkers in association with the clinical AD spectrum phenotype in a well-characterized AD cohort.

In this study, we analyzed cross-sectional asso-117 ciations of soluble plasma VCAM-1, ICAM-1, and 118 E-Selectin to baseline diagnosis and functional sever-119 ity staging in Alzheimer's Disease Neuroimaging 120 Initiative (ADNI) participants. ADNI is a highly 121 selective cohort that has been specifically enriched 122 for biomarker studies of AD-related cognitive decline 123 [63]. Utilizing the three bloodborne CAMs that 124 were available in ADNI plasma proteomics data, 125 we hypothesized that concomitantly higher plasma 126 CAM levels would correlate with severity of base-127 line diagnosis and functional impairment along the 128 prodromal-to-clinical AD dementia spectrum and 129 tested for modifying effects of Apolipoprotein E 130 (APOE) genotyping on functional impairment. Based 131 upon suggestions of interactive associations of CAMs 132 and AD pathology in recent experimental mod-133 els, we also hypothesized peripheral CAMs would 134 have a synergistic relationship with brain-based 135 AD biomarkers in influencing baseline functional 136 impairment, which we assessed using CSF biomark-137 ers of AD pathology and neurodegeneration in 138 a subset of subjects for whom those data were 139 available.

140

141 METHODS

142 Participants

Data used in this study were obtained from data of 143 the original ADNI cohort (http://adni.loni.usc.edu). 144 ADNI is a multi-site, longitudinal observational study 145 led by Principal Investigator Michael W. Weiner, 146 MD, that was initiated in 2004 as ADNI-1, and has 147 been extended by successive renewals to the cur-148 rent ADNI-3 cohort that was launched in 2016. The 149 primary goal of ADNI since its inception has been 150 to clarify the roles of imaging and other biomark-151 ers in AD clinical progression in order to validate 152 their use in AD clinical trials [63]. Written informed 153 consent was obtained for all participants, and prior 154 Institutional Review Board approval was obtained at 155 each participating institution. Up-to-date information 156 regarding ADNI can be found at http://www.adni-157 info.org. 158

159 Clinical and cognitive assessment

All subjects underwent an extensive clinical diagnostic evaluation, including basic mental status, neuropsychological, physical, and neurological examinations. A full description of assessment used can be found at http://adni.loni.usc.edu/methods/documents/

Dementia severity was graded by the Clinical Dementia Rating (CDR) Scale [64, 65], a measure that is widely employed in AD clinical care and research to quantify functional impairment. The CDR is derived through interview with patients and informants and consists of 6 domains (memory, orientation, judgment and problem solving, community affairs, home and hobbies, and personal care), each of which are rated on a 5-point scale (0, no impairment; 0.5, questionable impairment; 1, mild impairment; 2, moderate impairment; and 3, severe impairment). In clinical practice, the algorithm-generated global CDR score produces a total possible score of 0 to 3, denoting a global level of functional status from no impairment (global CDR 0) to severe impairment (global CDR 3) using the descriptors noted above for the individual domain box scores. The CDR sum of boxes score (CDR-SB), by contrast, utilizes a summary of the individual domain box scores and yields a total score of 0 to 18 (higher scores indicating greater impairment), and is frequently used in AD research given greater sensitivity in dementia staging and tracking of progression over time [66].

All participants were given diagnoses of cognitively normal (CN), mild cognitive impairment (MCI), or probable AD; for participants with MCI (global CDR score of 0.5), the inclusion criterion was an amnestic type (CDR memory domain box score of at least 0.5), to specifically ensure enrichment of the cohort with participants at high risk for conversion to AD. All AD patients satisfied NINCDS-ADRDA diagnostic criteria [67] for probable AD and had questionable to very mild dementia (global CDR score of 0.5 but considered borderline dementia) or mild (global CDR score of 1) dementia. Additionally, ADNI was specifically designed to minimize non-AD related risk factors for cognitive impairment or dementia, including vascular dementia. Inclusion criteria for ADNI-1 included a modified Hachinski ischemic score (MHIS) [68] of 4 or less to limit potential contributions from cerebrovascular disease; previously published baseline characteristics for ADNI-1 noted no significant difference between CN, MCI, and AD groups with respect to MHIS [69].

Biomarkers

ADNI proteomics data were collected for a subset of ADNI-1 (the original ADNI cohort) participants

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

who enrolled in this sub-study at baseline. The mul-212 tiplex panel was based upon Luminex immunoassay 213 technology and had been developed by Rules Based 214 Medicine (MyriadRBM) to measure a range of 215 inflammatory, metabolic, lipid, and other disease 216 relevant indices. A 190-analyte, plasma-based panel 217 of biomarkers previously reported to be related to 218 cell-signaling or disease processes such as AD, meta-219 bolic disorders, inflammation, cancer, and cardiovas-220 cular disease were analyzed through the Biomarkers 221 Consortium Project "Use of Targeted Multiplex 222 Proteomic Strategies to Identify Plasma-Based 223 Biomarkers in Alzheimer's Disease," 146 of 224 which met quality control standards. Biomarkers 225 of vascular endothelial dysfunction used in this 226 study included baseline peripheral blood-derived 227 VCAM-1, ICAM-1, and E-Selectin. Further details 228 regarding ADNI proteomics procedures can be 220 found in the data primer, "Biomarkers Consortium 230 Project: Use of Targeted Multiplex Proteomic 231 Strategies to Identify Plasma-Based Biomarkers in 232 Alzheimer's Disease" (http://adni.loni.usc.edu/wp-233 content/uploads/2010/12/BC-Plasma-Proteomics-234 Analysis-Plan.pdf). 235

A subset of ADNI participants also underwent a 236 lumbar puncture sub-study at the time of periph-237 eral blood collection at baseline. CSF specimens 238 for biomarkers were processed by the Biomarker 239 Core of ADNI at the Translational Research Lab-240 oratory, Department of Pathology & Laboratory 241 Medicine at the University of Pennsylvania Medi-242 cal School, under the direction of Drs. Leslie M. 243 Shaw and John Trojanowski. The Luminex multiplex 244 immunoassay platform was used for measurements 245 of amyloid- β , 42-residue peptide (A β_{42}), total tau 246 (T-tau), and phosphorylated tau (P-tau). Over 50 247 studies have demonstrated clinical sensitivity and 248 specificity for these biomarkers at greater than 249 80% each. Further details regarding ADNI CSF 250 Biomarker Core procedures can be found in the data 251 primer, "An Overview of the first 8 ADNI CSF 252 Batch Analyses" (http://adni.loni.usc.edu/methods/ 253 documents/). 254

APOE genotyping was performed at screening using established protocols, the details of which can be found at http://adni.loni.usc.edu/meth ods/documents/.

259 Statistical analyses

Distributions of peripheral blood levels of CAMs
 (VCAM-1, ICAM-1, and E-Selectin) and CSF

biomarkers (A β_{42} , T-tau, and P-tau) for the entire sample were assessed for normality. Using the Shapiro-Wilk test, all plasma CAM and CSF data were found to have non-normal distributions (all $p \le 0.005$). Thus, all plasma and CSF biomarkers were log-transformed for subsequent analysis, and then back transformed for graphical depictions.

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

A generalized linear model was used to assess VCAM-1, ICAM-1, and E-selectin concentrations (log normal) for each level of baseline diagnosis (CN, MCI, or AD). Using a generalized linear model (the GLIMMIX procedure), CDR-SB and the CDR memory box sub-scores (CDR-Mem) were modeled as binomial distributions by each dependent variable (VCAM-1, ICAM-1, and E-Selectin). We hypothesized that associations between the CDR-SB and each of the CAMs would be greater among those with the highest risk for AD, so we conducted additional analyses to evaluate effect modification by selected known risk factors for AD (e.g., APOE genotype: APOE ɛ4 non-carrier, APOE ɛ4 heterozygote, or APOE ε 4 homozygote), age, sex, and family history of AD). This would allow us to understand if the relationship between CDR-SB and each of the CAMS was being driven by factors other than our main independent variables. Interaction terms were tested for significance and included in the model when significant.

Using a generalized linear model (the GLIMMIX procedure), CDR-SB was modeled as a binomial distribution by the combination (fit plane) of peripheral plasma CAMs and CSF AD biomarkers (CSF A β_{42} , T-tau, P-tau, and P-tau/A β_{42} ratio, all log normal distributions). To understand if effects were interactive or additive, interaction terms were tested for significance and included if appropriate. Individual *p-values* of the dependent variables in the model were used to test the contribution of these variables. Interaction terms and correlations between CAMs and CSF biomarkers were assessed as a check against concerns over multicollinearity.

Familywise alpha was maintained at 0.05 using the Holm adjustment for multiple comparisons where appropriate (adjusted *p*-values are reported, unless otherwise stated). Significance level was set at 0.05 *a priori*. Classic sandwich estimation was used to adjust for any model misspecification. All statistical analyses were performed using SAS version 9.4 (The SAS Institute; Cary, NC).

313 **RESULTS**

314 Participants

Data were derived from the original ADNI cohort 315 (ADNI-1), which consisted of approximately 200 316 people with early AD, 400 people with MCI, and 317 200 CN older individuals. A cohort of 566 ADNI-318 1 participants (n=112 AD, n=396 MCI, n=58319 CN) for whom proteomics data were available were 320 included in the current study. Baseline demographic 321 data including age, sex, and education level (summa-322 rized in Table 1), showed expected diagnosis-related 323 characteristics with respect to CDR-SB and CDR-324 Mem scores. In our subset of the ADNI-1 cohort, we 325 observed the same discrepancy related to higher pro-326 portion of male participants in MCI versus CN and 327 AD groups that had previously been reported for the 328 entire ADNI-1 cohort [69]. There were no major dif-329 ferences in vascular risk (as measured by the MHIS) 330 between CN, MCI, and AD groups that would be 331 expected to confound our analyses (Table 1). 332

333 Baseline diagnosis

Plasma VCAM-1 levels for AD, MCI, and CN were 334 2.89 [2.87, 2.91], 2.86 [2.85, 2.87], and 2.84 [2.82, 335 2.87] ng/ml (mean [95% CI]), respectively (Fig. 1a). 336 Participants diagnosed with AD had significantly 337 higher mean VCAM-1 levels compared with the MCI 338 (p=0.0026) and CN (p=0.0028) diagnostic groups; 339 however, VCAM-1 levels in CN and MCI groups 340 were similar (CN versus MCI; p = 0.2687). Mean 341 ICAM-1 levels for AD, MCI, and CN were 2.04 [2.01, 342 2.07], 2.00 [1.98, 2.01] and 2.01 [1.97, 2.05] ng/ml. 343 Participants with AD had higher mean ICAM-1 con-344 centrations relative to those with MCI (p = 0.0189), 345 but not in comparison with the CN group (AD versus 346 CN; p = 0.4187) (Fig. 1b). ICAM-1 concentrations 347 were not significantly different in the CN versus MCI 348 group comparison (p = 0.5639). Mean E-Selectin lev-349 els for AD, MCI, and CN were 6.56 [6.11, 7.04], 6.58 350 [6.34, 6.84], and 6.79 [6.17, 7.47] ng/ml, respectively. 351 E-Selectin levels did not differ between patients with 352 AD, MCI, or CN diagnoses (p = 0.8280, Fig. 1c). 353

Functional status staging and CDR memory sub-score

Across the entire sample, baseline plasma VCAMl levels were associated with greater functional impairment stage at study entry as indicated by higher CDR-SB scores (p = 0.0157). Similarly, higher plasma VCAM-1 levels were associated with severity of memory impairment (as indicted by higher CDR-Mem sub-score) at baseline (p = 0.0071). Neither ICAM-1 (p = 0.0645 and p = 0.2489, respectively) nor E-selectin levels (p = 0.5700 and p = 0.6604, respectively) levels were associated with CDR-SB or CDR-Mem scores.

The effects of APOE genotype, age, sex, and family history

Severity of functional status (CDR-SB) modeled by VCAM-1 level and *APOE* genotype showed that *APOE* status modified the relationship of VCAM-1 to CDR-SB (Supplementary Fig. 1a). Although participants with higher CDR-SB scores also had higher VCAM-1 levels, regardless of *APOE4* status, we observed effect modification by genotype, such that *APOE* ε 4 heterozygotes (2.66 [2.39, 2.96]) and homozygotes (3.02 [2.47, 3.69]) (both *p* < 0.0001) had greater respective functional impairment scores at baseline (e.g., higher CDR-SB scores) than *APOE* ε 4 non-carriers (1.68 [1.50, 1.88]).

Age did not significantly influence the association of VCAM-1 with CDR-SB (Supplementary Figure 1b; slope not significantly different than 0, p=0.1395). Stratifying the sample by sex, we found a trend for female (slope significantly different from 0), but not male sex as a modifier of the relationship between VCAM-1 and CDR-SB (Supplementary Figure 1c; p=0.1888 and 0.0478 for men versus women, respectively). Family history of AD was also found to modify the relationship of VCAM-1 with CDR-SB. However, this was observed only for positive family history in both parents; this finding was significant before and after adjustment for variance (p=0.0148 and p=0.0444, respectively), but there were only 8 participants in this category (Table 1).

As only VCAM-1 was consistently correlated with clinical diagnosis and global functional status, the above analyses were not performed with respect to ICAM-1 or E-Selectin levels.

CSF AD biomarkers

About half of participants also underwent lumbar puncture by ADNI-1 protocol for collection of CSF samples at the baseline visit. For the following analyses, we included a subset of the original sample (n=351; n=102 AD, n=197 MCI, n=57 CN) with AD biomarker results in CSF (AB42, T-tau, P-tau, and

5

361 362 363

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

350

	I			
Characteristic, entire sample	CN	MCI	AD	
Demographics	n = 58	n = 396	n = 112	
Sex (% Female)	48.3	35.4	42.0	
Age, y (Median, [1 IQR, 3 IQR])	73.2 [71.1, 79.0]	75.1 [70.2, 80.4]	75.9 [69.4, 80.4]	
Education, y (Median, [1 IQR, 3 IQR])	16 [13, 18]	16 [14, 18]	16 [13, 18]	
AD family history	17.2/8.6	20.0/6.8	22.3/8.9	
(% maternal/paternal)				
APOE (% E4 Non-carrier/	92/8/0	46/42/12	32/47/21	
Heterozygote/ Homozygote)				
CDR-SB (Median, [1 IQR, 3 IQR])	0 [0, 0]	1.5 [1, 2]	4 [3.5, 5]	
CDR-Mem (Median, [1 IQR, 3 IQR])	0 [0, 0]	0.5 [0.5, 0.50]	1 [1, 1]	
MHIS (mean, SD, range)	0.64, 0.79, 0-3	0.62, 0.70, 0-4	0.66, 0.68, 0–3	
Characteristic, CSF subgroup	CN	MCI	AD	
Demographics	n = 57	n = 197	n = 102	
Sex (% Female)	47.4	33.0	42.2	
Age, y (Median, [1 IQR, 3 IQR])	73.1 [71.1, 78.6]	74.6 [70.1, 79.6]	75.9 [70.7, 80.5]	
Education, y (Median, [1 IQR, 3 IQR])	16 [13, 18]	16 [14, 18]	16 [13, 18]	
AD family history	17.5/8.8	24.3/8.1	22.6/9.8	
(% maternal/paternal)				
APOE (% E4 Non-carrier/	91/9/0	47/43/10	30/47/23	
Heterozygote/ Homozygote)				
CDR-SB (Median, [1 IQR, 3 IQR])	0 [0, 0]	1.5 [1, 2]	4 [3.5, 5]	
CDR-Mem (Median, [1 IQR, 3 IQR])	0 [0, 0]	0.5 [0.5, 0.5]	1 [1, 1]	
MHIS (mean, SD, range)	0.63, 0.79, 0-3	0.59, 0.77, 0-3	0.66, 0.70, 0-3	

 Table 1

 Data summary for ADNI-1 participants included in study

AD, Alzheimer's disease; ADNI, Alzheimer's Disease Neuroimaging Initiative; CDR-Mem, clinical dementia rating-memory subscale; CDR-SB, clinical dementia rating-sum of boxes; CN, cognitively normal; CSF, cerebrospinal fluid; E4, *APOE* £4 allele; 1 IQR, first interquartile range; 3 IQR, third interquartile range; MCI, mild cognitive impairment; MHIS, modified Hachinski ischemic score; SD, standard deviation.

Dependent Variable	CAMs	р	CSF biomarkers	р	DF
CDR-SB	VCAM-1	0.0022	Αβ ₄₂	< 0.0001	353
		0.0084	T-tau	< 0.0001	348
		0.0073	P-tau	< 0.0001	354
		0.0050	P-tau/Aβ ₄₂	< 0.0001	353
	ICAM-1	0.0713	Αβ ₄₂	< 0.0001	353
		0.0882	T-tau	< 0.0001	348
		0.0467	P-tau	< 0.0001	354
		0.0496	P-tau/A β_{42}	< 0.0001	353
	E-Selectin	0.2799	Αβ ₄₂	< 0.0001	353
		0.3539	T-tau	< 0.0001	348
		0.2462	P-tau	< 0.0001	354
		0.2400	P-tau/AB42	< 0.0001	353

Table 2 CDR-SB modeled (additive) by the combination of VCAM-1, ICAM-1, or E-Selectin and CSF biomarkers

 $A\beta_{42}$, amyloid- β 42-residue peptide; CDR-SB, clinical dementia rating-sum of boxes; DF, degrees of freedom; ICAM-1, intercellular adhesion molecule-1; P-tau, phosphorylated tau; T-tau, total tau; VCAM-1, vascular cell adhesion molecule-1.

P-tau/A β_{42} ratio, log normal distributions), as well 407 as measures of plasma VCAM-1, ICAM-1, and E-408 Selectin from concurrently collected blood samples. 409 We found that neither VCAM-1, ICAM-1, nor E-410 selectin levels across our sample at baseline were 411 significantly associated with any of the CSF biomark-412 ers (AB42, T-tau, or P-tau), or with P-tau/AB42 ratios 413 (data not shown). Moreover, contrary to our predic-414 tion, there was no evidence of interactive association 415

when CAMs were entered with CSF biomarkers into 416 the CDR-SB models individually (data not shown). 417 Instead, we observed significant additive relation-418 ships for VCAM-1 when included in CDR-SB 419 models with A β_{42} (p = 0.0022), T-tau (p = 0.0084), 420 P-tau (p = 0.0084), or P-tau/A β_{42} ratios (p = 0.005); 421 we found additive contributions of ICAM-1 to CDR-422 SB models that included either P-tau (p=0.0467) or 423 the P-tau/A β_{42} ratio (p = 0.0496) (Table 2).



Fig. 1. Associations of VCAM-1, ICAM-1, and E-Selectin with baseline diagnosis (figures in logarithmic scale). Dots with bars indicate average levels with confidence intervals. AD, Alzheimer's disease; ADNI; CN, cognitively normal; ICAM-1, intercellular adhesion molecule-1; MCI, mild cognitive impairment; ng/mL, nanograms per milliliter; VCAM-1, vascular cell adhesion molecule-1; *p < 0.05; **p < 0.005.

424 DISCUSSION

Current epidemiologic evidence suggests that 425 potentially modifiable midlife vascular risk factors 426 specifically influence risk for later-life, AD-ass-427 ociated cognitive decline. Despite this, a large knowl-428 edge gap still exists with respect to characterization 429 of the biological overlap of highly co-morbid vas-430 cular and AD pathologies in clinical AD. Given 431 recent emphasis on prevention-oriented strategies 432 in AD, delineating the nature of clinical associa-433 tions between vascular-mediated processes and AD 434 biomarkers is imperative. 435

In this study, we first sought to explore whether molecular indicators vascular endothelial dysfunction as indicated by soluble plasma CAM levels would reliably estimate baseline AD-related clinical diagnosis and/or functional impairment in a cohort that was purposefully designed for studying biomarkers in AD, in which contributions from potential confounders such as significant vascular cognitive impairment had been minimized. In our cross-sectional analyses of ADNI-1 data, we found that sample-wide baseline VCAM-1 and ICAM-1 (but not E-Selectin) levels had significant but modest associations with AD-associated diagnosis. We had

445

446

447

448

predicted that CAM levels would be lowest in CN 110 subjects and concomitantly higher in MCI and AD 450 groups, respectively, but found VCAM-1 and ICAM-451 1 to only be consistently higher in AD versus MCI 452 groups. As an extension of our initial hypothesis, we 453 had predicted that higher baseline peripheral CAMs 454 levels would also correlate with severity of functional 455 impairment staging and found that VCAM-1 (but not 456 ICAM-1 or E-Selectin) again had a significant but rel-457 atively modest relationship to CDR-SB scores across 458 the entire sample. 459

Our main finding was related to our prediction that 460 plasma CAMs and brain-based AD pathology would 461 act synergistically in contributing to clinical sever-462 ity of AD. Contrary to our initial expectation, we 463 found that plasma VCAM-1 and CSF AD biomarkers 464 acted independently and observed more robust addi-465 tive effects when added to any of the CSF biomarkers 466 in CDR-SB models. ICAM-1 was also observed to 467 have an additive effect in these models, but to a more 468 limited extent, and we observed no effects with E-469 Selectin in this model. 470

Our hypothesis that sensitive molecular indicators 471 of vascular endothelial dysfunction such as solu-472 ble plasma CAMs would indicate interactive rather 473 than independent associations of vascular and AD 474 pathologies was largely informed by recent experi-475 mental data. Peripheral immune cell trafficking into 476 the brain has been reported in rodent models to 477 occur with specific predilection for brain parenchyma 478 affected by deposition of A β [70, 71] and tau [72], 479 and one study reported high levels of both VCAM-480 1 and ICAM-1 expression in cerebral blood vessels 481 adjacent to AB plaques [71]. Moreover, AB has 482 been demonstrated to induce a CAM-mediated pro-483 inflammatory cascade in vascular endothelial cells 484 [73], blood-brain barrier (BBB) dysfunction via 485 endothelial and smooth muscle cell damage [74], and 486 vasoconstriction via free radicals [75]. Interestingly, 487 elevated pro-inflammatory CSF tumor necrosis fac-488 tor has been shown in a mouse model to drastically 489 increase β-site amyloid precursor protein-cleaving 490 enzyme 1 (BACE1) processing of CSF VCAM-1 491 from its membrane-bound to soluble form, a process 492 that was not prominent in healthy adult mice [76]. 493

Additionally, Yousef et. al. have demonstrated in a
detailed series of studies of aging mice that a) the aged
hippocampus expresses an inflammatory transcriptional profile that induces local microglial activation
that is spatially associated with focal VCAM-1 upregulation on the luminal side of the adjacent BBB; b)
soluble VCAM-1 is elevated in aged mouse (and

human) plasma, and aged plasma from mice and 501 humans induces VCAM-1 expression in cultured 502 brain endothelial cells and young mouse hippocampi; 503 and c) the effects of aged blood in this model 504 (including impaired cognition in a Barnes maze) 505 are mitigated by administration of an anti-VCAM-506 1 antibody or genetic ablation of VCAM-1 [58]. 507 The proinflammatory effects of the dialyzed, aged 508 plasma that was used for experiments were specif-509 ically found to not be related to soluble VCAM-1 510 itself, and authors surmised that pro-inflammatory 511 cytokine/chemokine signaling was likely responsi-512 ble for the effect [58]. Of note, they found no 513 increased expression of ICAM-1, E-Selectin, or P-514 Selectin at either mRNA or protein levels in their 515 model [58]. Yousef et al. also observed three distinct 516 populations of vascular endothelial cells, only two 517 of which expressed VCAM-1 together with either 518 pro-inflammatory genes or vascular remodeling and 519 Notch signaling markers [58]. It is interesting to note 520 parallels of these findings with an ApoE -/- mouse 521 model of atherosclerosis, where VCAM-1, ICAM-522 1, and PECAM were all shown to be differentially 523 expressed and localized in response to hypercholes-524 terolemia; however, only VCAM-1 upregulation 525 showed focal predilection for lesion-prone sites and 526 preceded atherosclerotic lesion formation [41]. 527

Human clinical studies to-date have consistently 528 demonstrated peripheral soluble CAMs to be elevated 529 in relation to various forms of dementia but have 530 yielded inconsistent results as to which CAMs are 531 most clinically relevant. For example, some groups 532 using different combinations of CAMs have shown 533 only VCAM-1 [49], ICAM-1 [51], or ICAM-1 and 534 PECAM-1 [50] to be elevated in AD compared with 535 controls, while Huang et al. found VCAM-1, ICAM-536 1, and E-Selectin to all be elevated in AD compared 537 with controls, but only VCAM-1 to be associated 538 with dementia severity [48]. With respect to cerebral 539 vascular disease, E-Selectin (but not VCAM-1) was 540 associated with severity of small vessel disease on 541 CT in patients with vascular dementia and AD in one 542 study [49]; however, Huang et. al. found VCAM-1 543 (but not ICAM-1 or E-Selectin) to be associated with 544 cerebral vascular dysfunction in AD patients as mea-545 sured by major tract-specific fractional anisotropy 546 quantification [48]. In two cross-sectional examina-547 tions of community-dwelling older adults, Tchalla 548 et al. found that elevated plasma VCAM-1 (but 549 not ICAM-1) was associated with cognitive impair-550 ment, decline in activities of daily living, slowed gait 551 speed, higher cerebral white matter hyperintensity 552

606

607

608

609

610

611

612

613

614

615

616

617

618

619

620

621

622

623

624

625

626

627

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647

648

649

650

651

652

653

volume on MRI, and cerebrovascular resistance as 553 measured by transcranial doppler [52, 53]. Another 554 group assessed an expanded panel of neuroinflamma-555 tory biomarkers including VCAM-1 and ICAM-1 in 556 serum (and CSF) for cross-sectional associations with 557 CSF AD biomarkers in normal community-dwelling 558 adults and patients with cognitive impairment, and 559 found that eight of the serum (including ICAM-1, 560 but not VCAM-1) biomarkers best predicted a CSF 561 AD profile defined by P-tau/A β_{42} ratio [55]. Lon-562 gitudinally, the Rotterdam study showed baseline 563 α1-antichymotrypsin, interleukin-6, and C-reactive 564 protein but not VCAM-1 or ICAM-1 to predict 565 future dementia risk [77], while Yoon et. al. reported 566 that soluble VCAM-1, ICAM-1, E-Selectin, and P-567 Selectin assessed longitudinally in healthy volunteers 568 all increased with age, but only elevated ICAM-1 pre-569 dicted poorer cognitive performance years later [32]. 570 To our knowledge, ours is the first study to assess 571 for interactive versus additive associations of blood-572 borne soluble CAMs and CSF AD biomarkers with 573 respect to AD diagnosis and dementia severity stag-574 ing in a highly characterized AD cohort. 575

There were several limitations of our study. These 576 included the cross-sectional design, multiple compar-577 isons, and the smaller number of participants with 578 CSF biomarkers relative to the overall cohort size. For 579 simplicity, we specifically chose to limit this initial 580 study to cross-sectional outcomes of AD-associated 581 diagnosis and functional impairment in order to test 582 our basic assumption that elevated CAMs would 583 generally serve as a useful biomarker of vascular con-584 tributions to the clinical AD phenotype. Although our 585 data suggest primacy of VCAM-1 among the CAMs 586 tested herein with respect to clinical AD, and par-587 allel rather than interactive associations of soluble 588 plasma VCAM-1 and ICAM-1 to CSF biomarkers 589 of AD-pathology and neurodegeneration, these data 590 should be interpreted with caution in the context 591 of findings from other groups that were generated 592 in studies with differing methodologies as outlined 593 above. It is also possible that available sample sizes 594 and/or the presence of outliers in ADNI data may have 595 influenced our findings related to between-group dif-596 ferences in ICAM-1 and E-Selectin. Additionally, if 597 membrane-bound VCAM-1 is in fact the key factor 598 in mediating vascular-AD pathological interactions 599 across the BBB as suggested by experimental stud-600 ies, the relative degrees to which individual CAMs are 601 expressed longitudinally, solubilized, and detectable 602 as blood-borne biomarkers in humans with chronic 603 risk factors remains unclear. It may also be that 604

use of blood-borne CAMs in isolation without proinflammatory chemokine/cytokine and/or imaging biomarkers is insufficient to fully capture clinically relevant aspects of vascular contributions to AD via mechanisms acting across the BBB.

Despite these limitations, our data lend weight to the growing literature suggesting that soluble bloodborne CAMs may serve as useful biomarkers for studying the biological overlap of vascular and AD pathologies in clinical AD. In order to validate these findings, future studies of CAMs in ADNI data will include assessment of associations with AD and vascular imaging biomarkers, correlations with longitudinal cognitive and functional data, and likely inclusion of CAMs in a more comprehensive panel of peripheral inflammation and BBB dysfunction. Future studies would also benefit from validation across multiple large, longitudinal cohorts using standardized methodologies.

ACKNOWLEDGMENTS

This work was supported by NIA # AG036535 and NIA #AG058648 (Daiello). The funding sources had no role in the design, conduct, or reporting of this study.

Data collection and sharing for this project was funded by the Alzheimer's Disease Neuroimaging Initiative (ADNI) (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Lumosity; Lundbeck; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support

ADNI clinical sites in Canada. Private sector con-654 tributions are facilitated by the Foundation for the 655 National Institutes of Health (http://www.fnih.org). 656 The grantee organization is the Northern Califor-657 nia Institute for Research and Education, and the 658 study is coordinated by the Alzheimer's Therapeutic 659 Research Institute at the University of Southern Cali-660 fornia. ADNI data are disseminated by the Laboratory 661 for NeuroImaging at the University of Southern 662 California. 663

Authors' disclosures available online (https:// www.j-alz.com/manuscript-disclosures/20-0759r2).

Prior Presentations: This work was presented, in part, at the Alzheimer's Association International Conference, Chicago, IL (July 2018 and July 2020).

669 SUPPLEMENTARY MATERIAL

The supplementary material is available in the electronic version of this article: https://dx.doi.org/ 10.3233/JAD-200759.

673 **REFERENCES**

- [1] Selkoe DJ, Hardy J (2016) The amyloid hypothesis of
 Alzheimer's disease at 25 years. *EMBO Mol Med* 8,
 595–608.
 - [2] de Bruijn RFAG, Bos MJ, Portegies MLP, Hofman A, Franco OH, Koudstaal PJ, Ikram MA (2015) The potential for prevention of dementia across two decades: The prospective, population-based Rotterdam Study. *BMC Med* 13, 132.
 - [3] Dar TA, Sheikh IA, Ganie SA, Ali R, Singh LR, Gan SH, Kamal MA, Zargar MA (2014) Molecular linkages between diabetes and Alzheimer's disease: Current scenario and future prospects. *CNS Neurol Disord Drug Targets* 13, 290–298.
 - [4] Mosconi L, Walters M, Sterling J, Quinn C, McHugh P, Andrews RE, Matthews DC, Ganzer C, Osorio RS, Isaacson RS, De Leon MJ, Convit A (2018) Lifestyle and vascular risk effects on MRI-based biomarkers of Alzheimer's disease: A cross-sectional study of middle-aged adults from the broader New York City area. *BMJ Open* 8, e019362.
 - [5] Arvanitakis Z, Capuano AW, Lamar M, Shah RC, Barnes LL, Bennett DA, Schneider JA (2018) Late-life blood pressure association with cerebrovascular and Alzheimer disease pathology. *Neurology* 91, e517–e525.
 - [6] Gilsanz P, Mayeda ER, Glymour MM, Quesenberry CP, Mungas DM, DeCarli C, Dean A, Whitmer RA (2017) Female sex, early-onset hypertension, and risk of dementia. *Neurology* 89, 1886–1893.
 - [7] McGrath ER, Beiser AS, DeCarli C, Plourde KL, Vasan RS, Greenberg SM, Seshadri S (2017) Blood pressure from mid- to late life and risk of incident dementia. *Neurology* 89, 2447–2454.
 - [8] De La Torre JC (2013) Vascular risk factors: A ticking time bomb to Alzheimer's disease. Am J Alzheimers Dis Other Demen 28, 551–559.

- [9] Kivipelto M, Helkala E, Laakso MP, Hänninen T, Hallikainen M, Alhainen K, Soininen H, Tuomilehto J, Nissien A (2001) Midlife vascular risk factors and Alzheimer's disease in later life: Longitudinal, population based study. *BMJ* 322, 1447–1451.
- [10] Lourida I, Hannon E, Littlejohns TJ, Langa KM, Hyppönen E, Kuzma E, Llewellyn DJ (2019) Association of lifestyle and genetic risk with incidence of dementia. *JAMA* 322, 430-437.
- [11] Gottesman RF, Schneider ALC, Zhou Y, Coresh J, Green E, Gupta N, Knopman DS, Mintz A, Rahmim A, Sharrett AR, Wagenknecht LE, Wong DF, Mosley TH (2017) Association between midlife vascular risk factors and estimated brain amyloid deposition. JAMA 317, 1443–1450.
- [12] Matthews FE, Arthur A, Barnes LE, Bond J, Jagger C, Robinson L, Brayne C, Medical Research Council Cognitive Function and Ageing Collaboration (2013) A two-decade comparison of prevalence of dementia in individuals aged 65 years and older from three geographical areas of England: Results of the Cognitive Function and Ageing Study I and II. *Lancet* 382, 1405–1412.
- [13] Attems J, Jellinger KA (2014) The overlap between vascular disease and Alzheimer's disease–lessons from pathology. *BMC Med* 12, 206.
- [14] Jagust W, Jack CR, Bennett DA, Blennow K, Haeberlein SB, Holtzman DM, Jessen F, Karlawish J, Liu E, Molinuevo JL, Montine T, Phelps C, Rankin KP, Rowe CC, Scheltens P, Siemers E, Sperling R (2019) "Alzheimer's disease" is neither "Alzheimer's clinical syndrome" nor "dementia". *Alzheimers Dement* 15, 153–157.
- Sweeney MD, Montagne A, Sagare AP, Nation DA, Schnei-[15] der LS, Chui HC, Harrington MG, Pa J, Law M, Wang DJJ, Jacobs RE, Doubal FN, Ramirez J, Black SE, Nedergaard M, Benveniste H, Dichgans M, Iadecola C, Love S, Bath PM, Markus HS, Salman RA, Allan SM, Quinn TJ, Kalaria RN, Werring DJ, Carare RO, Touyz RM, Williams SCR, Moskowitz MA, Katusic ZS, Lutz SE, Lazarov O, Minshall RD, Rehman J, Davis TP, Wellington CL, González HM, Yuan C, Lockhart SN, Hughes TM, Chen CLH, Sachdev P, O'Brien JT, Skoog I, Pantoni L, Gustafson DR, Biessels GJ, Wallin A, Smith EE, Mok V, Wong A, Passmore P, Barkof F, Muller M, Breteler MMB, Román GC, Hamel E, Seshadri S, Gottesman RF, van Buchem MA, Arvanitakis Z, Schneider JA, Drewes LR, Hachinski V, Finch CE, Toga AW, Wardlaw JM, Zlokovic BV (2019) Vascular dysfunction-The disregarded partner of Alzheimer's disease. Alzheimers Dement 15, 158-167.
- [16] Schneider JA, Arvanitakis Z, Bang W, Bennett DA (2007) Mixed brain pathologies account for most dementia cases in community-dwelling older persons. *Neurology* 69, 2197–2204.
- [17] Nelson PT, Dickson DW, Trojanowski JQ, Jack CR, Boyle PA, Arfanakis K, Rademakers R, Alafuzoff I, Attems J, Brayne C, Coyle-Gilchrist ITS, Chui HC, Fardo DW, Flanagan ME, Halliday G, Hokkanen SRK, Hunter S, Jicha GA, Katsumata Y, Kawas CH, Keene CD, Kovacs GG, Kukull WA, Levey AI, Makkinejad N, Montine TJ, Murayama S, Murray ME, Nag S, Rissman RA, Seeley WW, Sperling RA, White Iii CL, Yu L, Schneider JA (2019) Limbicpredominant age-related TDP-43 encephalopathy (LATE): Consensus working group report. *Brain* 74, 625–634.
- [18] Launer LJ, Petrovitch H, Ross GW, Markesbery W, White LR (2008) AD brain pathology: Vascular origins? Results from the HAAS autopsy study. *Neurobiol Aging* 29, 1587–15.

664

665

666

667

668

677

678

679

680

681

682

683

684

685

686

687

688

689

690

691

692

693

694

695

696

697

698

699

700

701

702

703

704

705

706

707

- [19] Jellinger KA (2007) The enigma of vascular cognitive disor der and vascular dementia. *Acta Neuropathol* 113, 349–388.
- 775 [20] Puzo C, Labriola C, Sugarman MA, Tripodis Y, Martin B, Palmisano JN, Steinberg EG, Stein TD, Kowall NW, 776 777 McKee AC, Mez J, Killiany RJ, Stern RA, Alosco ML (2019) Independent effects of white matter hyperintensities 778 on cognitive, neuropsychiatric, and functional decline: A 770 longitudinal investigation using the National Alzheimer's 780 781 Coordinating Center Uniform Data Set. Alzheimers Res Ther 11, 64. 782
- [21] Arvanitakis Z, Capuano AW, Leurgans SE, Bennett DA,
 Schneider JA (2016) Relation of cerebral vessel disease
 to Alzheimer's disease dementia and cognitive function in
 elderly people: A cross-sectional study. *Lancet Neurol* 15,
 934–943.
- [22] De La Torre JC (2000) Critically attained threshold of cerebral hypoperfusion: Can it cause Alzheimer's disease? *Ann N Y Acad Sci* 903, 424–436.
 - [23] Lo RY, Jagust WJ (2012) Vascular burden and Alzheimer disease pathologic progression. *Neurology* 79, 1349–1355.

792

796

797

798

799

- [24] Zlokovic BV (2011) Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci* 12, 723–38.
 - [25] Roseborough A, Ramirez J, Black SE, Edwards JD (2017) Associations between amyloid β and white matter hyperintensities: A systematic review. *Alzheimers Dement* 13, 1154–1167.
- [26] Koncz R, Sachdev PS (2018) Are the brain's vascular and
 Alzheimer pathologies additive or interactive? *Curr Opin Psychiatry* **31**, 147–152.
- Rabin JS, Schultz AP, Hedden T, Viswanathan A, Marshall [27] 803 GA, Kilpatrick E, Klein H, Buckley RF, Yang H-S, Prop-804 805 erzi M, Rao V, Kirn DR, Papp K V, Rentz DM, Johnson KA, Sperling RA, Chhatwal JP (2018) Interactive associations of 806 vascular risk and β-amyloid burden with cognitive decline 807 in clinically normal elderly individuals: Findings from 808 the Harvard Aging Brain Study. JAMA Neurol. 75, 1124-809 1131. 810
- [28] Banerjee G, Kim HJ, Fox Z, Jäger HR, Wilson D, Charidimou A, Na HK, Na DL, Seo SW, Werring DJ (2017)
 MRI-visible perivascular space location is associated with Alzheimer's disease independently of amyloid burden. *Brain* 140, 1107–1116.
- [29] Alosco ML, Sugarman MA, Besser LM, Tripodis Y, Martin
 B, Palmisano JN, Kowall NW, Au R, Mez J, DeCarli C, Stein
 TD, McKee AC, Killiany RJ, Stern RA (2018) A clinicopathological investigation of white matter hyperintensities
 and Alzheimer's disease neuropathology. *J Alzheimers Dis*63, 1347–1360.
- [30] Soldan A, Pettigrew C, Zhu Y, Wang M, Moghekar A,
 Gottesman RF, Singh B, Martinez O, Fletcher E, DeCarli
 C, Albert M, BIOCARD Research Team (2020) White matter hyperintensities and CSF Alzheimer disease biomarkers
 in preclinical Alzheimer disease. *Neurology* 94, e950-e960.
- [31] Price DT, Loscalzo J (1999) Cellular adhesion moleculesand atherogenesis. *Am J Med* **107**, 85–97.
- [32] Yoon CY, Steffen LM, Gross MD, Launer LJ, Odegaard A,
 Reiner A, Sanchez O, Yaffe K, Sidney S, Jacobs DR (2017)
 Circulating cellular adhesion molecules and cognitive func tion: The coronary artery risk development in young adults
 study. *Front Cardiovasc Med* 4, 37.
- [33] Wittchen ES (2009) Endothelial signaling in paracellular
 and transcellular leukocyte transmigration. *Front Biosci* (*Landmark Ed*) 14, 2522-2545.

- [34] Muller WA (2009) Mechanisms of transendothelial migration of leukocytes. *Circ Res* 105, 223–230.
- [35] Muller WA (2015) The regulation of transendothelial migration: New knowledge and new questions. *Cardiovasc Res* 107, 310–320.
- [36] Yousef H, Czupalla C, Lee D, Burke A, Chen M, Zandstra J, Berber E, Lehallier B, Mathur V, Nair R, Bonanno L, Merkel T, Schwaninger M, Quake S, Butcher E, Wyss-Coray T (2019) Aged blood inhibits hippocampal neurogenesis and activates microglia through VCAM1 at the blood-brain barrier. *Nat Med* 25, 988–1000.
- [37] Kallmann BA, Hummel V, Lindenlaub T, Ruprecht K, Toyka KV, Rieckmann P (2000) Cytokine-induced modulation of cellular adhesion to human cerebral endothelial cells is mediated by soluble vascular cell adhesion molecule-1. *Brain* 123 (Pt 4), 687–697.
- [38] Deanfield JE, Halcox JP, Rabelink TJ (2007) Endothelial function and dysfunction: Testing and clinical relevance. *Circulation* 115, 1285–1295.
- [39] Kong D-H, Kim YK, Kim MR, Jang JH, Lee S (2018) Emerging roles of vascular cell adhesion molecule-1 (VCAM-1) in immunological disorders and cancer. *Int J Mol Sci* 19, 13–17.
- [40] Ewers M, Mielke MM, Hampel H (2010) Blood-based biomarkers of microvascular pathology in Alzheimer's disease. *Exp Gerontol* 45, 75–9.
- [41] Nakashima Y, Raines EW, Plump AS, Breslow JL, Ross R (1998) Upregulation of VCAM-1 and ICAM-1 at atherosclerosis-prone sites on the endothelium in the ApoE-deficient mouse. *Arterioscler Thromb Vasc Biol* 18, 842–851.
- [42] Hwang SJ, Ballantyne CM, Sharrett AR, Smith LC, Davis CE, Gotto AM, Boerwinkle E (1997) Circulating adhesion molecules VCAM-1, ICAM-1, and E-selectin in carotid atherosclerosis and incident coronary heart disease cases: The Atherosclerosis Risk In Communities (ARIC) study. *Circulation* 96, 4219–4225.
- [43] Blann AD, Lip GYH (2000) Cell adhesion molecules in cardiovascular disease and its risk factors What can soluble levels tell us? *J Clin Endocrinol Metab* **85**, 1745–1747.
- [44] Qiu S, Cai X, Liu J, Yang B, Zügel M, Steinacker JM, Sun Z, Schumann U (2019) Association between circulating cell adhesion molecules and risk of type 2 diabetes: A meta-analysis. *Atherosclerosis* 287, 147–154.
- [45] Fassbender K, Bertsch T, Mielke O, Mühlhauser F, Hennerici M (1999) Adhesion molecules in cerebrovascular diseases. Evidence for an inflammatory endothelial activation in cerebral large- and small-vessel disease. *Stroke* 30, 1647–1650.
- [46] Rieckmann P, Nünke K, Burchhardt M, Albrecht M, Wiltfang J, Ulrich M, Felgenhauer K (1993) Soluble intercellular adhesion molecule-1 in cerebrospinal fluid: An indicator for the inflammatory impairment of the blood-cerebrospinal fluid barrier. *J Neuroimmunol* 47, 133–140.
- [47] Sharief MK, Noori MA, Ciardi M, Cirelli A, Thompson EJ (1993) Increased levels of circulating ICAM-1 in serum and cerebrospinal fluid of patients with active multiple sclerosis. Correlation with TNF- α and blood-brain barrier damage. J Neuroimmunol **43**, 15–21.
- [48] Huang C-W, Tsai M-H, Chen N-C, Chen W-H, Lu Y-T, Lui C-C, Chang Y-T, Chang W-N, Chang AYW, Chang C-C (2015) Clinical significance of circulating vascular cell adhesion molecule-1 to white matter disintegrity in Alzheimer's dementia. *Thromb Haemost* **114**, 1230–1240.

837

838

839

840

841

842

843

844

845

846

847

848

849

850

851

852

853

854

855

856

857

858

859

860

861

862

863

864

865

866

867

868

869

870

871

872

873

874

875

876

877

878

879

880

881

882

883

884

885

886

887

888

889

890

891

892

893

894

895

896

897

898

899

- [49] Zuliani G, Cavalieri M, Galvani M, Passaro A, Munari MR, Bosi C, Zurlo A, Fellin R (2008) Markers of endothelial dysfunction in older subjects with late onset Alzheimer's disease or vascular dementia. J Neurol Sci 272, 164– 170.
- [50] Nielsen HM, Londos E, Minthon L, Janciauskiene SM (2007) Soluble adhesion molecules and angiotensinconverting enzyme in dementia. *Neurobiol Dis* 26, 27–35.
- [51] Rentzos M, Michalopoulou M, Nikolaou C, Cambouri C, Rombos A, Dimitrakopoulos A, Kapaki E, Vassilopoulos D (2004) Serum levels of soluble intercellular adhesion molecule-1 and soluble endothelial leukocyte adhesion molecule-1 in Alzheimer's disease. J Geriatr Psychiatry Neurol 17, 225–231.
- [52] Tchalla AE, Wellenius GA, Sorond FA, Gagnon M, Iloputaife I, Travison TG, Dantoine T, Lipsitz LA (2017) Elevated soluble vascular cell adhesion molecule-1 is associated with cerebrovascular resistance and cognitive function. J Gerontol A Biol Sci Med Sci 72, 560–566.
- [53] Tchalla AE, Wellenius GA, Travison TG, Gagnon M, Iloputaife I, Dantoine T, Sorond FA, Lipsitz LA (2015) Circulating vascular cell adhesion molecule-1 is associated with cerebral blood flow dysregulation, mobility impairment, and falls in older adults. *Hypertension* 66, 340–346.
- [54] Janelidze S, Mattsson N, Stomrud E, Lindberg O, Palmqvist S, Zetterberg H, Blennow K, Hansson O (2018) CSF biomarkers of neuroinflammation and cerebrovascular dysfunction in early Alzheimer disease. *Neurology* 91, e867–e877.
- [55] Popp J, Oikonomidi A, Tautvydaité D, Dayon L, Bacher M, Migliavacca E, Henry H, Kirkland R, Severin I, Wojcik J, Bowman GL (2017) Markers of neuroinflammation associated with Alzheimer's disease pathology in older adults. *Brain Behav Immun* 62, 203–211.
- [56] Gupta VB, Hone E, Pedrini S, Doecke J, O'Bryant S, James I, Bush AI, Rowe CC, Villemagne VL, Ames D, Masters CL, Martins RN, AIBL Research Group (2017) Altered levels of blood proteins in Alzheimer's disease longitudinal study: Results from Australian Imaging Biomarkers Lifestyle Study of Ageing cohort. *Alzheimers Dement (Amst)* 8, 60–72.
- [57] Hochstrasser T, Weiss E, Marksteiner J, Humpel C (2010) Soluble cell adhesion molecules in monocytes of Alzheimer's disease and mild cognitive impairment. *Exp Gerontol* 45, 70–74.
- [58] Yousef H, Czupalla CJ, Lee D, Chen MB, Burke AN, Zera KA, Zandstra J, Berber E, Lehallier B, Mathur V, Nair R V, Bonanno LN, Yang AC, Peterson T, Hadeiba H, Merkel T, Körbelin J, Schwaninger M, Buckwalter MS, Quake SR, Butcher EC, Wyss-Coray T (2019) Aged blood impairs hippocampal neural precursor activity and activates microglia via brain endothelial cell VCAM1. *Nat Med* 25, 988–1000.
- [59] Li G, Shofer JB, Petrie EC, Yu CE, Wilkinson CW,
 Figlewicz DP, Shutes-David A, Zhang J, Montine TJ,
 Raskind MA, Quinn JF, Galasko DR, Peskind ER (2017)
 Cerebrospinal fluid biomarkers for Alzheimer's and vascular disease vary by age, gender, and APOE genotype in
 cognitively normal adults. *Alzheimers Res Ther* 9, 48.
- [60] Markus HS, Hunt B, Palmer K, Enzinger C, Schmidt H, Schmidt R (2005) Markers of endothelial and hemostatic activation and progression of cerebral white matter
 hyperintensities: Longitudinal results of the Austrian Stroke
 Prevention Study. *Stroke* 36, 1410–1414.

- [61] Verbeek MM, Otte-Höller I, Westphal JR, Wesseling P, Ruiter DJ, de Waal RM (1994) Accumulation of intercellular adhesion molecule-1 in senile plaques in brain tissue of patients with Alzheimer's disease. Am J Pathol 144, 104–116.
- [62] Li G, Xiong K, Korff A, Pan C, Quinn JF, Galasko DR, Liu C, Montine TJ, Peskind ER, Zhang J (2015) Increased CSF E-selectin in clinical Alzheimer's disease without altered CSF Aβ42 and tau. J Alzheimers Dis 47, 883–887.
- [63] Veitch DP, Weiner MW, Aisen PS, Beckett LA, Cairns NJ, Green RC, Harvey D, Jack CR, Jagust W, Morris JC, Petersen RC, Saykin AJ, Shaw LM, Toga AW, Trojanowski JQ, Alzheimer's Disease Neuroimaging Initiative (2019) Understanding disease progression and improving Alzheimer's disease clinical trials: Recent highlights from the Alzheimer's Disease Neuroimaging Initiative. *Alzheimers Dement* 15, 106–152.
- [64] Morris JC (1993) The Clinical Dementia Rating (CDR): Current version and scoring rules. *Neurology* 43, 2412–2414.
- [65] Hughes CP, Berg L, Danziger WL, Coben LA, Martin RL (1982) A new clinical scale for the staging of dementia. Br J Psychiatry 140, 566–572.
- [66] O'Bryant SE, Waring SC, Cullum CM, Hall J, Lacritz L, Massman PJ, Lupo PJ, Reisch JS, Doody R, Texas Alzheimer's Research Consortium (2008) Staging dementia using Clinical Dementia Rating Scale Sum of Boxes scores: A Texas Alzheimer's research consortium study. *Arch Neurol* 65, 1091–1095.
- [67] McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM (1984) Clinical diagnosis of Alzheimer's disease: Report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology* 34, 939–944.
- [68] Hachinski VC, Iliff LD, Zilhka E, Du Boulay GH, McAllister VL, Marshall J, Russell RW, Symon L (1975) Cerebral blood flow in dementia. *Arch Neurol* 32, 632–637.
- [69] Petersen RC, Aisen PS, Beckett LA, Donohue MC, Gamst AC, Harvey DJ, Jack CR, Jagust WJ, Shaw LM, Toga AW, Trojanowski JQ, Weiner MW (2010) Alzheimer's Disease Neuroimaging Initiative (ADNI): Clinical characterization. *Neurology* 74, 201–209.
 - [70] Buckwalter MS, Coleman BS, Buttini M, Barbour R, Schenk D, Games D, Seubert P, Wyss-Coray T (2006) Increased T cell recruitment to the CNS after amyloid beta 1-42 immunization in Alzheimer's mice overproducing transforming growth factor-beta 1. *J Neurosci* 26, 11437–11441.
 - [71] Fisher Y, Nemirovsky A, Baron R, Monsonego A (2011) Dendritic cells regulate amyloid-β-specific T-cell entry into the brain: The role of perivascular amyloid-β. J Alzheimers Dis 27, 99–111.
 - [72] Majerova P, Michalicova A, Cente M, Hanes J, Vegh J, Kittel A, Kosikova N, Cigankova V, Mihaljevic S, Jadhav S, Kovac A (2019) Trafficking of immune cells across the bloodbrain barrier is modulated by neurofibrillary pathology in tauopathies. *PLoS One* 14, e0217216.
 - [73] Suo Z, Tan J, Placzek A, Crawford F, Fang C, Mullan M (1998) Alzheimer's β-amyloid peptides induce inflammatory cascade in human vascular cells: The roles of cytokines and CD40. *Brain Res* 807, 110–117.
 - [74] Rhodin JA, Thomas TN, Clark L, Garces A, Bryant M (2003) In vivo cerebrovascular actions of amyloid β-

12

901

902 903

904

905

906

907

908

909

910

911

012

913

914

915

916

917

918

919

920

921

922

923

924

925

926

927

928

929

930

931

932

933

934

935

936

937

938

939

940

941

942

943

944

945

946

947

948

949

950

951

952

peptides and the protective effect of conjugated estrogens. *J Alzheimers Dis* **5**, 275–286.

[75] Thomas T, Thomas G, McLendon C, Sutton T, Mullan
 M (1996) β-Amyloid-mediated vasoactivity and vascular
 endothelial damage. *Nature* 380, 168–171.

1029

1030

- 1034 [76] Voytyuk I, Mueller SA, Herber J, Snellinx A, Moechars D,
 1035 van Loo G, Lichtenthaler SF, De Strooper B (2018) BACE2
 1036 distribution in major brain cell types and identification of
 1037 novel substrates. *Life Sci Alliance* 1, e201800026.
- [77] Engelhart MJ, Geerlings MI, Meijer J, Kiliaan A, Ruitenberg A, van Swieten JC, Stijnen T, Hofman A, Witteman JCM, Breteler MMB (2004) Inflammatory proteins in plasma and the risk of dementia: The rotterdam study. *Arch Neurol* 61, 668–672.